

Variation in Caffeine Concentration in Single Coffee Beans

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ABSTRACT: Twenty-eight coffee samples from around the world were tested for caffeine levels to develop near-infrared reflectance spectroscopy (NIRS) calibrations for whole and ground coffee. Twenty-five individual beans from five of those coffees were used to develop a NIRS calibration for caffeine concentration in single beans. An international standard high-performance liquid chromatography method was used to analyze for caffeine content. Coffee is a legal stimulant and possesses a number of health properties. However, there is variation in the level of caffeine in brewed coffee and other caffeinated beverages. Being able to sort beans on the basis of caffeine concentration will improve quality control in the level of caffeine in those beverages. The range in caffeine concentration was from 0.01 mg/g (decaffeinated coffee) to 19.9 mg/g (Italian coffee). The majority of coffees were around 10.0–12.0 mg/g. The NIRS results showed r^2 values for bulk unground and ground coffees were >0.90 with standard errors <2 mg/g. For the single-bean calibration the r^2 values were between 0.85 and 0.93 with standard errors of cross validation of 0.8–1.6 mg/g depending upon calibration. The results showed it was possible to develop NIRS calibrations to estimate the caffeine concentration of individual coffee beans. One application of this calibration could be sorting beans on caffeine concentration to provide greater quality control for high-end markets. Furthermore, bean sorting may open new markets for novel coffee products.

KEYWORDS: caffeine, coffee bean, ground coffee, bean sorting, near infrared spectroscopy

■ INTRODUCTION

Coffee is one of the most-consumed beverages in the world with aroma and taste being the main sensory characteristics. The dominant commercial species of coffee are Arabica (*Coffea arabica* L.) and Robusta (*Coffea canephora* Pierre), and most commercial coffee beverages are produced from these roasted beans or blends of these two. However, there are differences in quality and sensory profiles between these species, and suitable analytical methods and trained taste panels are required to quantify these differences.

In regard to quantifying chemical components, such as lipids, protein, or caffeine concentration, expensive and time-consuming wet chemistry methods are required. However, near-infrared reflectance spectroscopy (NIRS) has been used as a high-throughput and cheap surrogate for quantification of quality (including caffeine concentration)^{1–7} and sensory parameters.⁸ The differences detected in the chemical composition within and between Arabica and Robusta varieties can be useful for quality classification purposes, and NIRS has been used successfully in this context.¹ In addition, NIRS has also been used for quantification of green bean quality^{9,10} and to understand genetic, environmental, and processing effects on quality.¹¹

In most biological studies, a bulk sample is tested to obtain a representative mean value, although there is little information on the variation within the sample and the impact of the variation on the result. Several studies in cereal crops investigated the evaluation of single grains to ascertain the variation within a sample for a range of attributes using NIRS.^{12–24} To date, there has been no report of assessment of single coffee beans by NIRS. However, single coffee beans have been tested for specific purposes including optimizing a wet

chemistry method for caffeine concentration²⁵ and identifying breeding lines for reducing caffeine concentration.^{2,26}

The aim of our study was to develop NIRS calibrations for caffeine concentration in bulk unground and ground coffees as well as single coffee beans using a high-performance liquid chromatography (HPLC) method as the reference method to quantify caffeine levels. The NIRS calibration in bulk unground and ground coffees would provide a rapid, predictive method of caffeine concentration. A NIRS calibration on single beans would provide the opportunity to understand the potential variability in coffee samples, for example, blends, or sort beans on the basis of caffeine concentration to further quantify quality and potentially develop new products (e.g., naturally low-caffeine coffee).

■ MATERIALS AND METHODS

Twenty-eight roasted Arabica and blended whole bean coffees, purchased from retail outlets, from a number of countries were used. Most of the coffees were Arabica type as per the information on the packet. Unfortunately, some of packets lacked specific or complete information on the origin or blend of the coffee. Table 1 shows the types and origins of coffees.

Ground Coffee. A 20 g subsample from each coffee was ground in a standard electric coffee grinder for NIR scanning and HPLC analysis. The grinder was cleaned with a fine brush between coffees.

FT-NIR. Three forms of coffees (bulk unground, ground, and single beans) were scanned in a Fourier transform (FT) NIR multipurpose analyzer instrument (Bruker, Germany) from 4000 to 12000 at 8 cm resolution. For bulk unground and ground coffees, the 97 mm cup on

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Table 1. Caffeine Concentration of Samples Collected

| coffee | brand ^a | type ^a | origin ^a | caffeine (mg/g) |
|--------|---|-------------------|---|-----------------|
| 1 | Brasilia Coffee Italian Espresso | blend | Italy | 17.7 |
| 2 | Brasilia Coffee Supa Cream | blend | Italy | 12.7 |
| 3 | Oxfam Aus World Blend Organic Espresso | Arabica | Africa/Latin American/Asia Pacific | 12.5 |
| 4 | Vittoria Espresso Coffee | blend | Italy | 12.2 |
| 5 | Caffe Aurora Medaglia Doro | blend | Italy | 19.9 |
| 6 | Monjava | blend | Central America | 12.4 |
| 7 | Dormans Breakfast (MR ^b) | blend | Kenya | 11.7 |
| 8 | Dormans Espresso (DR ^c) | blend | Kenya | 11.8 |
| 9 | Dormans AA Blue mountain (MR) | Arabica | Kenya | 11.4 |
| 10 | Masai (MR) | Arabica | Kenya | 10.8 |
| 11 | CTM Lush | blend | Colombia/Nicarigua/Timor/New Guinea | 10.7 |
| 12 | CTM House | blend | Brazil/Colombia/Sumatra/India | 12.3 |
| 13 | CTM Revive | blend | Colombia/Costa Rica/Indonesia/New Guinea/Brazil | 11.6 |
| 14 | CTM Jamocha | blend | South America/Central America/Africa/East India | 12.7 |
| 15 | CTM Mimic (decaf) | blend | Colombia/New Guinea | 0.01 |
| 16 | CTM Single Origin | blend | Rwanda | 10.6 |
| 17 | Woolworths Select Espresso | Arabica | | 12.2 |
| 18 | Gloria Jean's Coffee Smooth Classic Blend | Arabica | Central and South America | 12.1 |
| 19 | North Queensland Gold | Arabica | Australia | 11.6 |
| 20 | PNG Hand Roast | Arabica | Papua New Guinea | 12.2 |
| 21 | Brazil Hand Roast | Arabica | Brazil | 12.8 |
| 22 | Colombia Hand Roast | Arabica | Colombia | 12.8 |
| 23 | Doisaket Coffee Brand | Arabica | Thailand | 11.7 |
| 24 | Native Organic | Arabica | Brazil | 10.5 |
| 25 | Green Cauldron | Arabica | Australia | 10.7 |
| 26 | Café One Italian No. 5 | Arabica | Italy | 11.8 |
| 27 | Spar Espresso (SRR ^d) | Arabica | South Africa | 11.8 |
| 28 | Café de Chamarel L'Ile Maurice | | Europe | 11.7 |

^aAs per information on sample packet. ^bMR, medium roast. ^cDR, dark roast. ^dSR, strong rich roast.

the rotating sphere was used, where 64 scans of the rotating cup were averaged into a single spectrum. Coffees were scanned in duplicate.

Five coffees were selected for single-bean analysis (coffees 5, 7, 11, 15, and 24 in Table 1). Four samples were blends that would provide a broad range within each coffee. A single Arabica was also included. Twenty-five individual beans were randomly selected from each of these five coffees and scanned in a 19 mm glass vial using the microsphere attachment. A chrome insert was placed on top of each bean to prevent light escaping, as per Bruker recommendations. The wavenumber range and resolution were the same as for the bulk unground and ground coffees. After scanning, each bean was sealed in a plastic bag and stored in the freezer prior to grinding for caffeine analysis.

Opus (V 7.0) was used for calibration development with all pretreatments selected in the Optimise process to find optimal calibration math treatment and factors. Savistky–Golay, with first and second derivatives, was used with 4, 4 gap, and smoothing. Individual spectra files were exported in data point table format (Opus software), loaded into Excel, and then imported into Unscrambler [V 9.8,

(Camo, Sweden)] for calibration development. The calibrations developed from these two multivariate software programs were compared using the Fearn²⁷ calculation, which tests for significant differences ($P < 0.05$) between the standard deviation of prediction errors of predicted values (from cross-validation) using both packages.

Caffeine Analysis. For the ground coffees, 2.000 g of coffee was weighed out on a three decimal place analytical balance. For single beans, each bean was ground in the coffee grinder and recovered. The recovered coffee was weighed and used for caffeine extraction.

The weighed coffee was dissolved in boiling water, mixed well, and kept heated for 10 min with constant stirring. The coffee solution was quickly cooled in iced water and then equilibrated at room temperature. The coffee solution was made up to 100 mL in a volumetric flask, mixed well, and then filtered through a no. 42 filter paper. Five milliliters of the filtrate was further filtered through a 0.45 μm mesh nylon membrane and diluted 1 in 100 mL in a volumetric flask for HPLC analysis.

A Waters HPLC system was used for caffeine quantification. Caffeine (see Figure 1 for structure) was separated in an Alltech

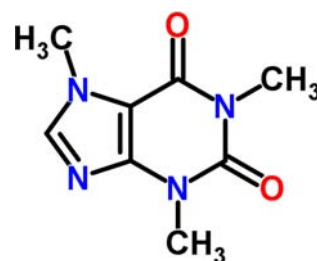


Figure 1. Structure of caffeine. Extracted from <http://www.chemspider.com/Chemical-Structure.2424.html>.

Prevail C18, 150 mm \times 4.6 mm, 5 μm , column with a guard column. Column temperature was 30 $^{\circ}\text{C}$ and injection volume, 10 μL . The mobile phase was 40% methanol and 0.5% phosphoric acid. The final pressure in the instrument was 119–122 bar with the detector set at 254 nm. Caffeine was identified by matching peak retention time against a caffeine standard and quantified by plotting against the standard curve of prepared caffeine standards at 1, 5, 10, and 20 mg/mL.

RESULTS

Range in Caffeine between Coffees. Of the 28 coffees collected in this study, one (no. 15) was a decaffeinated coffee, whereas all others were caffeinated. The caffeine concentration of the decaffeinated coffee was 0.01 mg/g (Table 1). The range in caffeine concentration for the other 27 samples was from 10.6 and 19.9 mg/g (Table 1). Most coffees were between 10.6 and 12.8 mg/g, whereas coffees 1 and 5 were much higher (17.7 and 19.9 mg/g, respectively). These two high-caffeine coffees were obtained from Italy and, as per the information on the packet, were identified as blends.

Range in Caffeine within Selected Coffees for Single-Bean Analysis. Five of the 28 samples were selected to investigate variation in caffeine concentration within each sample. Table 2 shows the HPLC caffeine concentrations for the 25 beans from the 5 coffees. For all coffees used in the single-bean analysis, the calculated average of the 25 individual beans was slightly less than the result for the coffee when tested as a bulk ground coffee (Table 2). However, all five of these coffees had individual kernels well above the bulk ground average. For the decaffeinated coffee, all beans contained trace amounts of caffeine, whereas the high-caffeine coffee had eight individual beans above 20 mg/g, with one kernel at 25.3 mg/g.

Table 2. Variation in Caffeine Concentration (Milligrams per Gram) in 25 Single Beans from 5 Selected Coffees

| bean | sample | | | | |
|----------------------------|--------|------|------|-----|------|
| | 5 | 7 | 11 | 15 | 24 |
| 1 | 10.1 | 11.2 | 9.9 | 1.2 | 10.4 |
| 2 | 16.8 | 10.1 | 11.3 | 0.7 | 7.4 |
| 3 | 16.5 | 9.3 | 8.4 | 0.7 | 9.5 |
| 4 | 15.8 | 13.6 | 8.9 | 0.9 | 10.1 |
| 5 | 21.7 | 13.2 | 10.0 | 0.7 | 12.0 |
| 6 | 14.7 | 14.1 | 13.0 | 0.6 | 11.8 |
| 7 | 22.7 | 12.4 | 10.3 | 1.0 | 8.6 |
| 8 | 17.9 | 13.9 | 9.0 | 0.5 | 10.1 |
| 9 | 19.4 | 17 | 11.0 | 1.2 | 8.3 |
| 10 | 20.2 | 10.1 | 10.1 | 1.2 | 10.1 |
| 11 | 18.9 | 8.6 | 8.9 | 0.6 | 9.7 |
| 12 | 15.9 | 10.7 | 7.7 | 0.6 | 9.7 |
| 13 | 20.2 | 12.3 | 9.9 | 0.6 | 9.4 |
| 14 | 18.2 | 10.8 | 8.5 | 0.3 | 9.5 |
| 15 | 23.7 | 11.2 | 10.5 | 0.5 | 11.0 |
| 16 | 20.1 | 14.4 | 8.3 | 0.8 | 10.5 |
| 17 | 15.4 | 12 | 11.2 | 0.9 | 12.2 |
| 18 | 25.3 | 11 | 10.9 | 0.6 | 9.3 |
| 19 | 18 | 11.4 | 10.5 | 0.9 | 9.0 |
| 20 | 20.1 | 10.6 | 9.6 | 0.8 | 11.0 |
| 21 | 16.1 | 10 | 7.9 | 1.1 | 11.0 |
| 22 | 15.3 | 10 | 9.6 | 0.6 | 11.0 |
| 23 | 16.7 | 11.8 | 8.9 | 0.6 | 9.8 |
| 24 | 15.4 | 6.7 | 7.9 | 0.6 | 7.2 |
| 25 | 18.2 | 10.5 | 9.1 | 0.6 | 11.2 |
| calcd mean | 18.1 | 11.5 | 9.7 | 0.8 | 10.0 |
| median | 18.0 | 11.2 | 9.6 | 0.7 | 10.1 |
| SD | 3.2 | 2.8 | 1.2 | 0.3 | 1.3 |
| max | 25.3 | 14.4 | 13.0 | 1.2 | 12.2 |
| min | 14.7 | 6.7 | 7.7 | 0.6 | 7.2 |
| ground coffee ^a | 19.9 | 11.7 | 10.7 | 0.0 | 10.5 |

^aCaffeine concentration of bulk ground coffee for each of these coffees.

The distribution plots of caffeine concentration of each of the individual five coffees is shown in Figure 2.

Figure 3 shows the distribution plots for the weight of beans used for calibration development. A normal distribution was seen in bean weights for caffeinated and decaffeinated beans. Panels a and b of Figure 4 show the correlation plots between caffeinated beans and decaffeinated beans, respectively, when compared to bean weights. There was a positive relationship in caffeine between caffeinated beans and bean weights ($R^2 = 0.31$). There was a similar positive relationship between bean weight and caffeine concentration in decaffeinated beans ($R^2 = 0.34$). These relationships indicated around 30% of bean weight is associated with caffeine concentration, which would suggest around 70% of the caffeine concentration is not explained by weight. Hence, selecting for weight would not assist in calibration development if the goal was to select for heavier or lighter beans.

NIR Calibrations for Bulk Unground and Ground Coffees. For all calibrations developed, either a standard normal variation (SNV) first derivative or multiplicative scatter correction (MSC) first derivative was selected as best. All calibrations for bulk unground and ground beans had greater than eight factors. The Opus calibration for bulk unground and ground coffees resulted in high coefficients of determination of

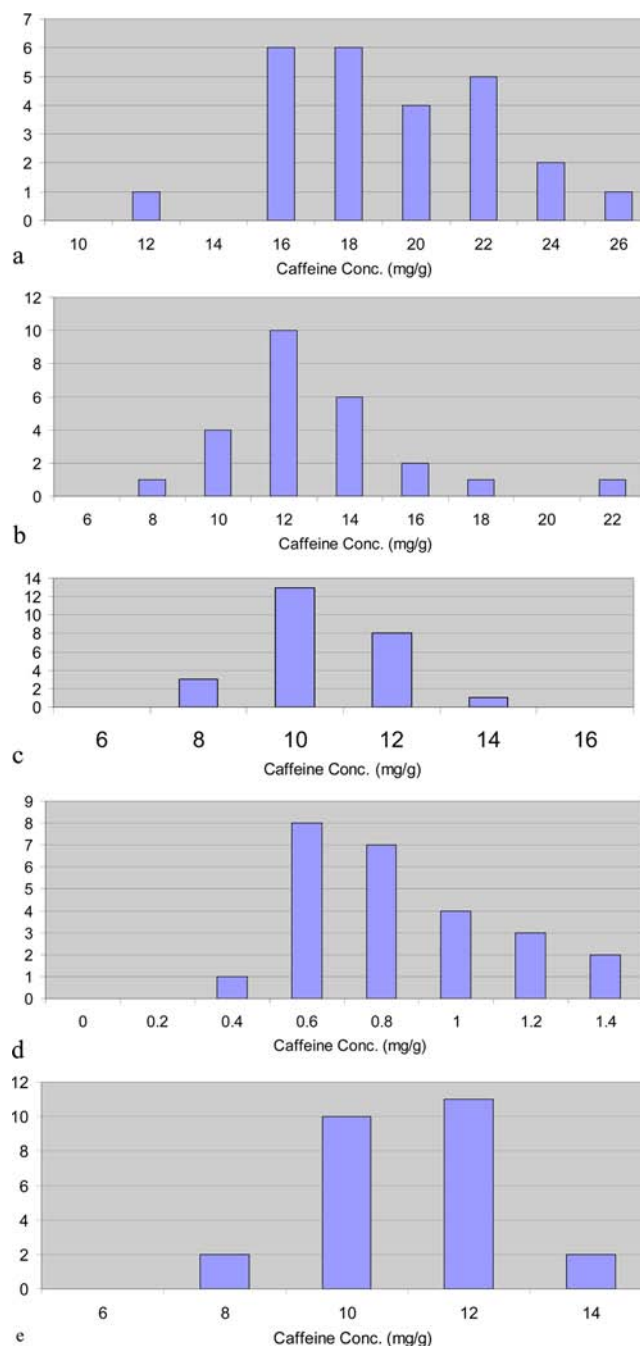


Figure 2. Distribution of caffeine between single beans of five samples: 5 (a); 7 (b); 11 (c); 15 (d); 24 (e).

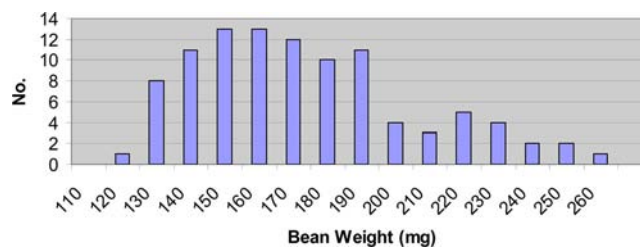


Figure 3. Distribution of bean weights used in NIRS calibration development.

0.93 and 0.95, respectively (Table 3), whereas in Unscrambler the r^2 values were 0.92 and 0.95, respectively. The r^2 values for

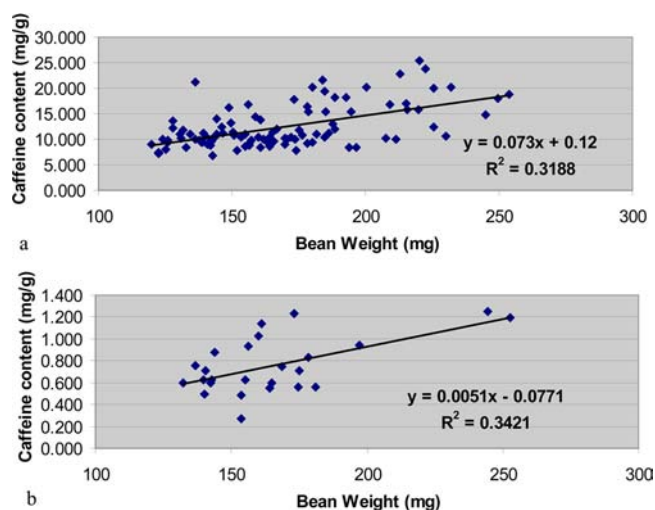


Figure 4. Correlation plot between bean weight and caffeine concentration for use in NIR calibration development for (a) caffeinated coffees and (b) decaffeinated coffee beans.

internal cross-validation were ≥ 0.92 for bulk unground or ground coffees. The NIRS correlation plots for the whole bulk unground and ground coffees are shown in Figures 4a and 5b, respectively.

The inclusion of a single decaffeinated coffee (coffee 15), with a zero level of caffeine well below the next coffee at 10.5 mg/g could have a major influence on the overall regression. Hence, the calibrations were repeated with the decaffeinated coffee excluded. The cross-validation r^2 values were again > 0.90 for unground and ground coffees using both multivariate analysis programs.

All standard error of calibration (SEC) results for whole bulk unground and ground coffees were < 1.2 and < 1.3 mg/g, respectively, by both modeling packages (Table 3). The SECs for the calibrations excluding the decaffeinated coffee showed a slight improvement.

Both multivariate analysis packages allowed for internal cross-validation. The internal standard error of cross-validation (SECV) for both unground and ground coffee was < 1.3 mg/g

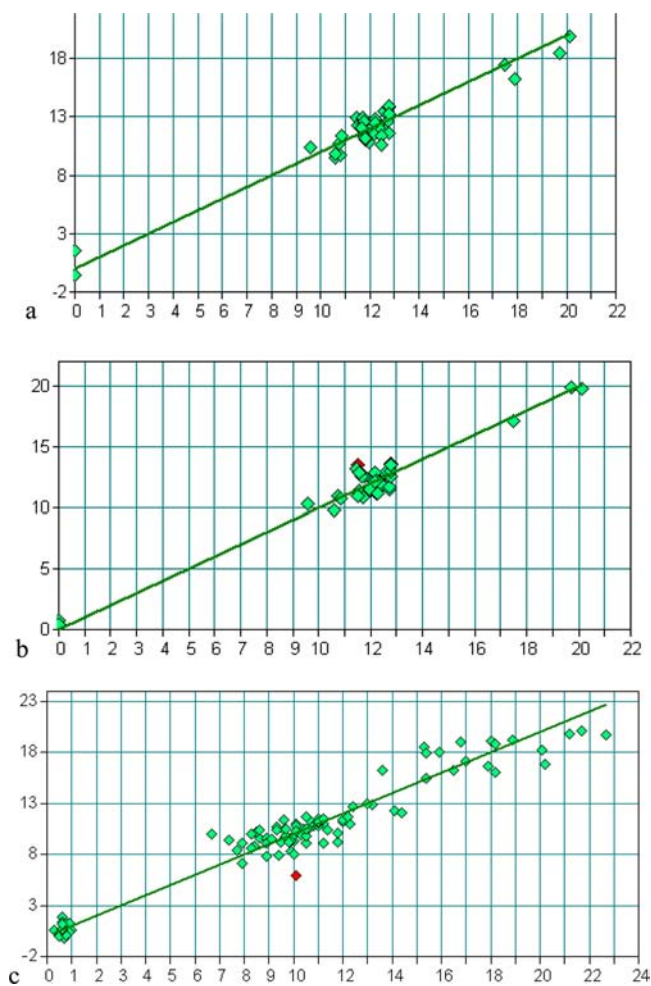


Figure 5. Correlation plots for whole (a), ground (b), and (c) single-bean coffees [actual (X axis) versus predicted (Y axis)].

(Table 3). The SECV decreased slightly when the decaffeinated coffee was excluded from the calibration set.

A ratio of standard error prediction to standard deviation (RPD) is used to indicate the potential application value of the

Table 3. Statistics for NIR Calibrations for Unground Bulk, Ground, and Single-Bean Coffees^a

| sample type | no. | R^2 | SEC | R^2_{CV} | SECV | RPD_{CV} | factors | bias | treatment (smoothing) |
|--------------------------------|-----|--------|-----|------------|------|------------|---------|--------|-----------------------|
| Opus calibrations | | | | | | | | | |
| bulk | 28 | 0.9325 | 0.9 | 0.9255 | 1.0 | 3.65 | 8 | 0.003 | 2dSNV (9) |
| ground | 28 | 0.9504 | 0.8 | 0.9333 | 0.9 | 4.10 | 5 | 0.001 | 2dSNV (9) |
| single bean | 124 | 0.9510 | 1.4 | 0.9197 | 1.7 | 3.53 | 10 | 0.005 | 1dSNV (13) |
| bulk less decaff coffee | 27 | 0.9311 | 0.9 | 0.9178 | 1.0 | 3.55 | 8 | 0.003 | 2dSNV (9) |
| ground less decaff coffee | 27 | 0.9434 | 0.8 | 0.9199 | 0.9 | 4.25 | 5 | 0.001 | 2dSNV (9) |
| single-bean less decaff coffee | 99 | 0.9304 | 1.1 | 0.8749 | 1.5 | 2.83 | 9 | 0.02 | 1dMSC (13) |
| Unscrambler calibrations | | | | | | | | | |
| bulk | 28 | 0.9225 | 0.9 | 0.9255 | 1.0 | 3.65 | 8 | 0.003 | 2dSNV |
| ground | 28 | 0.9511 | 0.8 | 0.9343 | 0.9 | 4.10 | 5 | 0.001 | 2dSNV |
| single bean | 122 | 0.9491 | 1.3 | 0.9228 | 1.6 | 3.55 | 8 | -0.013 | 1dSNV |
| bulk less decaff coffee | 27 | 0.9301 | 0.9 | 0.9098 | 1.0 | 3.52 | 8 | 0.003 | 2dSNV |
| ground less decaff coffee | 27 | 0.9407 | 0.8 | 0.9124 | 0.9 | 4.21 | 5 | 0.001 | 2dSNV |
| single-bean less decaff coffee | 97 | 0.9275 | 1.2 | 0.8599 | 1.6 | 2.75 | 8 | -0.02 | 1dSNV |

^aNo., number of samples used in the calibration; R^2 , coefficient of determination of calibration set; SEC, standard error of calibration; R^2_{CV} , coefficient of determination of cross-validation; SECV, standard error of cross-validation; RPD_{CV} , ratio of SD to standard error of prediction using the cross-validation.

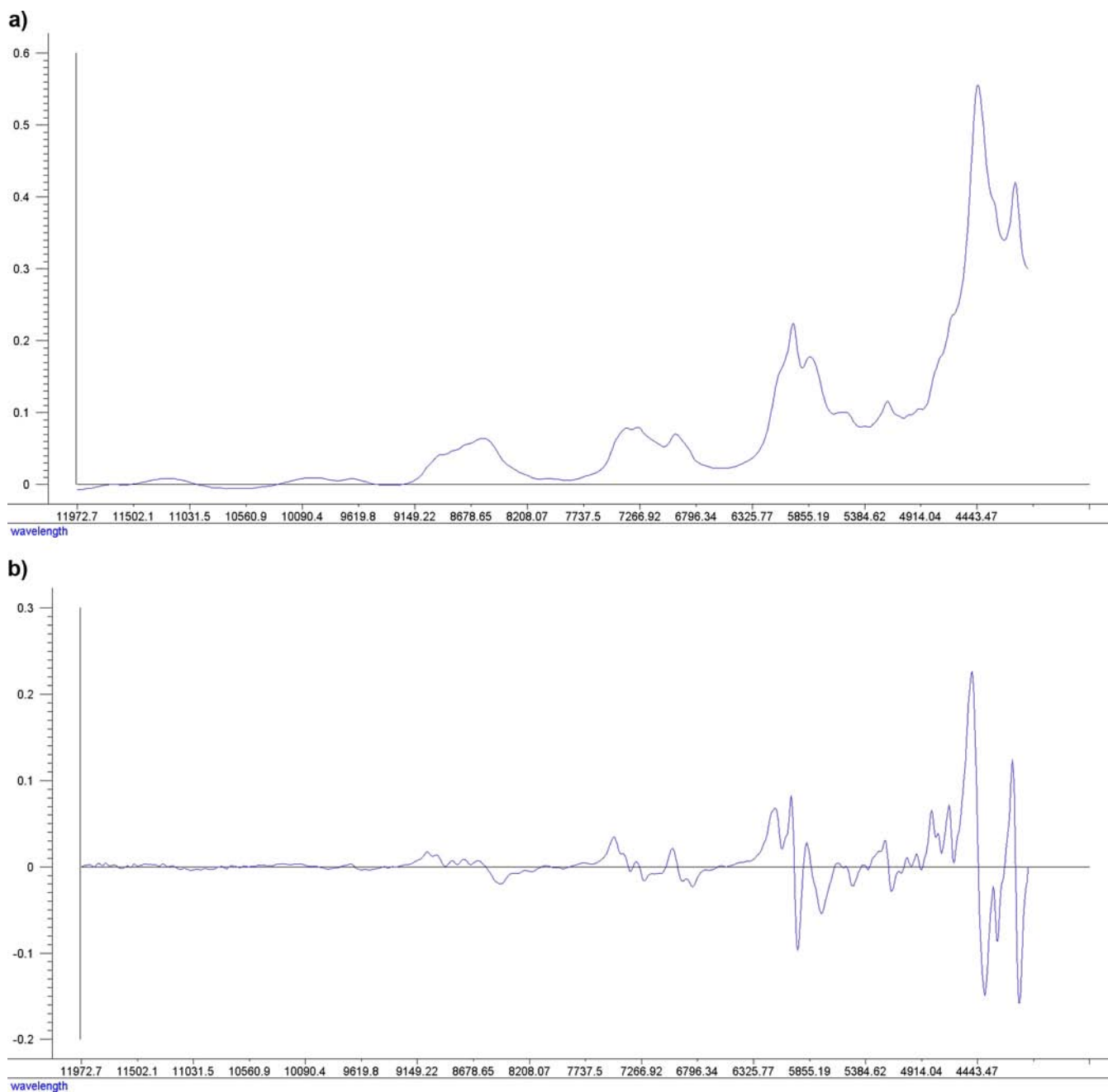


Figure 6. Pure caffeine standard: (a) raw spectra; (b) first-derivative SNV treatment.

calibration.²⁸ For RPD values >2.5 , the calibrations would be suitable for screening lines in breeding programs. For values >5 , the calibration would be useful in quality control. For the unground and ground calibrations using both multivariate programs, the RPDs were calculated at >4.0 for unground and ground coffees (Table 3), suggesting reasonably good calibrations for both product types. This was also the case when the decaffeinated coffee was excluded.

The bias for each calibration was also <0.1 , which is acceptable on the basis of the suggestion of Shenk, that is, bias $< 0.6 \times \text{SEC}$.

Fearn's Statistic. Fearn proposed a calculation to compare if standard deviations of prediction errors and bias between two different calibrations were significantly different when compared to reference values at $P < 0.10$, $P < 0.05$, or $P < 0.01$

levels. Using the Fearn calculation, updated by Reid and Guthrie, there was no significant difference ($P < 0.05$) between the standard deviations of prediction errors and bias for the calibration developed using the two multivariate packages for unground and ground coffee calibrations when the predicted values were compared to the reference values.

NIR Calibrations for Single Beans. The calibrations for the single beans produced high $r^2 > 0.94$ with both multivariate packages with the decaffeinated coffee included. When the decaffeinated coffee was excluded, the Opus program gave a $r^2 > 0.9304$, whereas the Unscrambler package gave a $r^2 = 0.9275$. The r^2 values for cross-validation were >0.91 from both programs when using all five coffees. However, when the decaffeinated coffee was excluded, the r^2 values were considerably lower (Table 3).

For single beans, the SECs and SECVs were similar to those for the unground and ground coffee calibrations. The $RPD_{(CV)}$ values were also similar to those of the unground and ground coffee RPDs with the calibrations with all five coffees included being higher than the RPDs for calibrations with the decaffeinated coffee excluded.

The Fearn statistic showed no significant difference ($P < 0.05$) for standard deviation of prediction errors and bias for predicted values from both multivariate packages when compared to the reference values.

Loading Plots/Wavelengths Associations. The raw spectra and first-derivative SNV spectra for the pure caffeine are shown in Figure 6, panels a and b, respectively. The structure of caffeine is shown in Figure 1, with obvious C=C, C=N, O=C—NR, and CH₃ bonds. These combination bonds are strongly associated with regions beyond 5000 cm⁻¹ (2200 nm). The Opus loading plots from the calibrations for bulk unground and ground coffees and single beans are shown in Figure 7, panels a, b, and c, respectively. These loading plots show a number of strong associations in regions associated with proteins and oils. In particular, there are a number of regions associated with NH amino acid groups and urea and C=O associated with oil groups.

DISCUSSION

The development of a NIRS calibration to predict caffeine content in dried coffee has been reported previously and using more samples than in this study.^{11,29} However, those studies used a relatively small number of samples but expanded the calibration set by either having samples from multiple locations or by collecting samples from different processing techniques, respectively. Our calibration used only 28 samples but included 2 samples from Australia, which is the first time caffeine content has been reported for Australian-grown coffee. These samples were within the average range of caffeine when compared to international samples.

The novel aspect of our research was the development of a single-bean NIRS calibration to predict caffeine concentration. We believe this to be the first report for a NIR calibration to predict caffeine concentration in single coffee beans. Even with the five different coffees used for the NIRS calibration, there was considerable variation in caffeine concentration. The variation in caffeine concentration was not dependent upon bean weight as only around 30% of caffeine concentration was correlated to bean weight, which indicated selecting for bean weight would not enhance selecting for caffeine concentration.

Our results for unground bulk and ground bulk coffee for caffeine concentration were similar to those of other researchers, who reported r^2 values >0.9 , with SECV or SEP around 1–2 mg/g. The potential application values of unground and ground bulk calibrations were similar, based on the RPD values, despite the low number of samples in our calibrations and the use of only a cross-validation approach. The ground coffee calibrations were the best models from both multivariate packages on the basis of the reduced and more uniform particle size. However, in the practical sense, the potential uses of NIR for any coffee trait would be of more value for the prediction and sorting of unground beans. As part of a quality control program at packaging facilities where beans are ground and undergo vacuum-seal packaging, then calibrations for ground coffee could be used.

The calibrations developed used a number of spectral regions associated with the major bonds in caffeine (C=O, C=C,

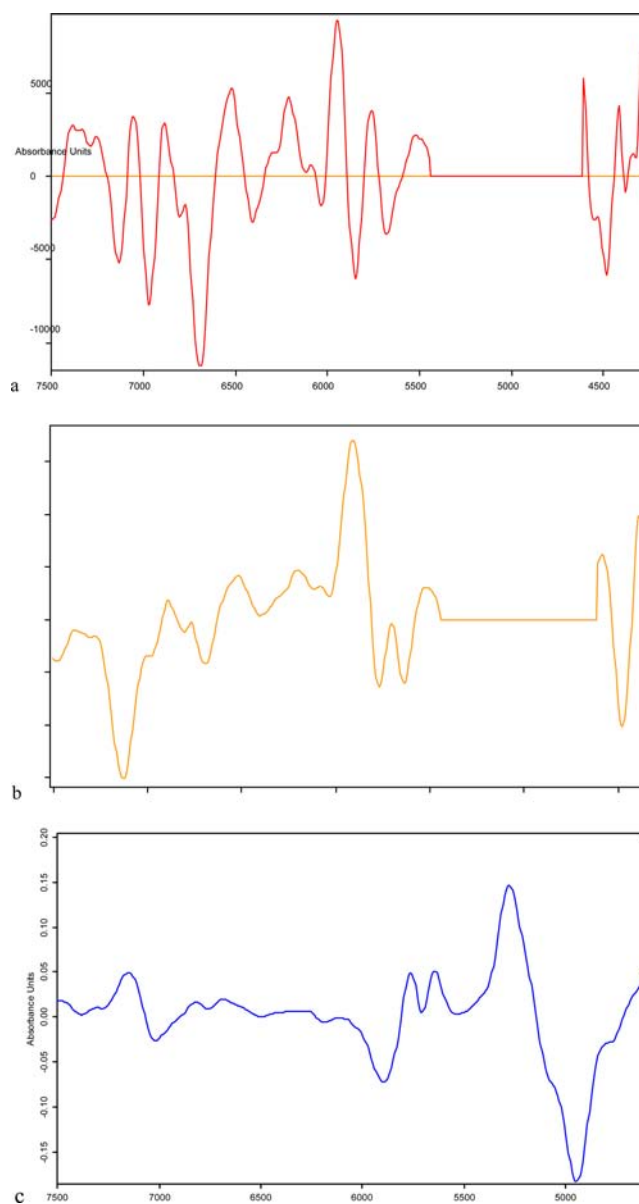


Figure 7. Loading plots for (a) bulk unground, (b) ground, and (c) single beans.

O=C—NR). These molecular bonds are also in protein and oil, which make up around 30% in coffee, so it could be expected to see these regions used in the calibration. In the spectra for pure caffeine, these regions also appeared as major peaks on a 1d spectrum. There are a number of regions identified that were associated with urea O=C—((NH₂)₂). The C=N bonds have also been identified in a study of hydrogen cyanide (HCN) in forage sorghum.³⁰

In our study, we compared two multivariate programs, one being the software of the FT-NIR instrument and the other being a commercial package that can use any type of multivariate data. There were some differences in the final calibrations developed using each package. However, as a real world application, using the software dedicated to operating the instrument is a better option. Exporting data into dedicated multivariate packages provides the scope to understand how good a calibration could be.

The development of a NIR calibration to predict caffeine concentration in single coffee beans has shown the potential to

understand the variation in pure Arabica coffees or blended coffees. In addition, to produce a decaffeinated coffee, the caffeine is removed from beans usually by chemical extraction. Using a nondestructive technique such as NIRS could assist processors in knowing the in situ concentration of caffeine in individual beans, which could then in turn help optimize the decaffeination process. Alternatively, this tool could be used to identify and sort naturally low (or high) caffeine beans to develop new coffee products and open new markets.

Although caffeine may not contribute to the major flavor (bitterness) of coffee, consumers purchase coffee with some anticipation of a stimulating effect from caffeine. On the basis of our results, it would be possible to build a calibration to sort beans on caffeine concentration. However, current commercial NIR sorting technology is quite limited in applications and costly, and further research and development are required.

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Notes

The authors declare no competing financial interest.

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REFERENCES

- (1) Downey, G.; Boussion, J. Authentication of coffee bean variety by near-infrared reflectance spectroscopy of dried extract. *J. Sci. Food Agric.* **1996**, *71*, 41–49.
- (2) Gous, P.; Martin, A.; Lawson, W.; Kelly, A.; Fox, G.; Sutherland, M. QTL associated with barley (*Hordeum vulgare*) feed quality traits measured through in situ digestion. *Euphytica* **2012**, *185*, 37–45.
- (3) Casal, S.; Oliveira, M.; Alves, M.; Ferreira, M. A. Discriminate analysis of roasted coffee varieties for trigonelline, nicotinic acid, and caffeine content. *J. Agric. Food Chem.* **2000**, *48*, 3420–3424.
- (4) Tripathi, S.; Mishra, H. A rapid FT-NIR method for estimation of aflatoxin B(1) in red chili powder. *Food Control* **2009**, *20*, 840–846.
- (5) Tallada, J.; Wicklow, D.; Pearson, T.; Armstrong, P. R. Detection of fungus-infected corn kernels using near-infrared reflectance spectroscopy and colour imaging. *Trans. ASABE* **2011**, *54*, 1151–1158.
- (6) McGoverin, C.; Engelbrecht, P.; Geladi, P.; Manley, M. Characterisation of non-viable whole barley, wheat and sorghum grains using near-infrared hyperspectral data and chemometrics. *Anal. Bioanal. Chem.* **2011**, *401*, 2283–2289.
- (7) Salgo, A.; Gergely, S.; Juhasz, R. Characterizing the maturation and germination processes in wheat by NIR methods. *Using Cereal Science and Technology for the Benefit of Consumers. Proceedings of the 12th International ICC Cereal and Bread Congress*, Harrogate, UK, May 23–26, 2004.
- (8) Ribeiro, J.; Ferreira, M.; Salva, T. Chemometric models for the quantitative descriptive sensory analysis of Arabica coffee beverages using near infrared spectroscopy. *Talanta* **2011**, *83*, 1352–1358.
- (9) Bertrand, B.; Etienne, H.; Lashermes, P.; Guyot, B.; Davrieux, F. Can near-infrared reflectance of green coffee be used to detect introgression in *Coffea arabica* cultivars? *J. Sci. Food Agric.* **2005**, *85*, 955–962.
- (10) Santos, J.; Sarraguca, M.; Rangel, A.; Lopes, J. Evaluation of green coffee beans quality using near infrared spectroscopy: a quantitative approach. *Food Chem.* **2012**, *135*, 1828–1835.
- (11) Posada, H.; Ferrand, M.; Davrieux, F.; Lashermes, P.; Bertrand, B. Near infrared spectral signature and their stability across environments. *22nd International Conference on Coffee Science*, ASIC 2008, Campinas, SP, Brazil, Sept 14–19, 2008.
- (12) Delwiche, S. Measurement of single-kernel wheat hardness using near-Infrared transmittance. *Trans. ASAE* **1993**, *36*, 1431–1437.
- (13) Delwiche, S. Single wheat kernel analysis by near-infrared transmittance protein content. *Cereal Chem.* **1995**, *72*, 11–16.
- (14) Angelino, S.; VanLaarhoven, H.; VanWesterop, J.; Broekhuijse, B.; Mocking, H. Total nitrogen content in single kernel malting barley samples. *J. Inst. Brew.* **1997**, *103*, 41–46.
- (15) Home, S.; Wilhelmson, A.; Tammissola, J.; Husman, J. Natural variation among barley kernels. *J. Am. Soc. Brew. Chem.* **1997**, *55*, 47–51.
- (16) Dowell, F. Automated color classification of single wheat kernels using visible and near-infrared reflectance. *Cereal Chem.* **1998**, *75*, 142–144.
- (17) Delwiche, S.; Hruschka, W. Protein content of bulk wheat from near-infrared reflectance of individual kernels. *Cereal Chem.* **2000**, *77*, 86–88.
- (18) Jiang, H.; Zhu, Y.; Wei, L.; Dai, J.; Song, T.; Yan, Y.; Chen, S. Analysis of protein, starch and oil content of single intact kernels by near infrared reflectance spectroscopy (NIRS) in maize (*Zea mays* L.). *Plant Breed.* **2007**, *126*, 492–497.
- (19) Pearson, T.; Brabec, D.; Dogan, H. Improved discrimination of soft and hard white wheat using SKCS and imaging parameters. *Sensing Instrum. Food Qual. Saf.* **2009**, 89–99.
- (20) Tallada, J.; Palacios-Rojas, N.; Armstrong, P. Prediction of maize seed attributes using a rapid single kernel near infrared instrument. *J. Cereal Sci.* **2009**, *50*, 381–387.
- (21) Fox, G.; Sweeney, N.; Hocroft, D. Development of a single kernel NIR barley protein calibration and assessment of variation in protein on grain quality. *J. Inst. Brew.* **2011**, *117*, 582–586.
- (22) Peiris, K.; Dowell, F. Determining weight and moisture properties of sound and *Fusarium*-damaged single wheat kernels by near-infrared spectroscopy. *Cereal Chem.* **2011**, *88*, 45–50.
- (23) Williams, P.; Geladi, P.; Fox, G.; Manley, M. Maize kernel hardness classification by near infrared (NIR) hyperspectral imaging and multivariate data analysis. *Anal. Chim. Acta* **2009**, *653*, 121–130.
- (24) Williams, P.; Manley, M.; Fox, G.; Geladi, P. Indirect detection of *Fusarium verticillioides* in maize (*Zea mays* L.) kernels by near infrared hyperspectral imaging. *J. Near Infrared Spec.* **2010**, *18*, 49–58.
- (25) Bonnlander, B.; Lonzarich, V.; Liverani, F. S. Caffeine determination in single beans by LC-MS/MS DAD. *21st International Conference on Coffee Science*, Montpellier, France, Sept 11–15, 2006.
- (26) Baumann, T.; Sondahl, M.; Waldhauser, S.; Kretschmar, J. Non-destructive analysis of natural variability in bean caffeine content of Laurina coffee. *Phytochemistry* **1998**, *49*, 1569–1573.
- (27) Reid, D.; Guthrie, J. Comparing two competing calibrations for a given data set using Fearn's criteria. Version 2.0. *NIR Spectrum* **2004**, *2*, 7.
- (28) Williams, P.; Sobering, D. Comparison of commercial near infrared transmittance and reflectance instruments for analysis of whole grains and seeds. *J. Near Infrared Spectrosc.* **1993**, *1*, 25–32.
- (29) Esteban-Diez, I.; Gonzalez-Saiz, J.; Pizarro, C. Prediction of roasting colour and other quality parameters of roasted coffee samples by near infrared spectroscopy. A feasibility study. *J. Near Infrared Spectrosc.* **2004**, *12*, 287–297.
- (30) Fox, G. P.; O'Donnell, N. H.; Stewart, P. N.; Gleadow, R. M. Estimating hydrogen cyanide in forage sorghum (*Sorghum bicolor*) by near-infrared spectroscopy. *J. Agric. Food Chem.* **2012**, *60*, 6183–6187.